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# Glucose metabolism-related genes with clinicopathological characteristics and prognosis of breast cancer: an analysis based on TCGA database

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**Abstract: Objective** To investigate the relationship between the expression of *LDHA*, *SLC16A1* and *SLC16A3* genes and pathologic features and prognosis in breast cancer. **Methods** Tissue samples from 1 060 breast cancer patients in The Cancer Genome Atlas (TCGA) were obtained. The association of *LDHA*, *SLC16A1* and *SLC16A3* gene expressions with clinicopathological features and prognosis of breast cancer were analyzed. Survival curve was drawn by Kaplan-Meier survival analysis, and univariable and multivariable survival prognoses were analyzed by Cox proportional hazard regression model. **Results** *LDHA* expression was associated with distant metastasis (M stage) ( $\chi^2$ =5.560, P=0.018), estrogen receptor (ER) expression ( $\chi^2$ =8.532, P=0.003), and human epidermal growth factor receptor 2 (HER-2) expression ( $\chi^2$ =4.418, P=0.036); *SLC16A1* expression correlated with age ( $\chi^2$ =8.040, P=0.005), ER expression ( $\chi^2$ =17.428, P<0.01), and progesterone receptor (PR) expression ( $\chi^2$ =5.486,  $\rho$ =0.019). *SLC16A3* expression correlated with ER expression ( $\chi^2$ =22.447,  $\rho$ <0.01) and PR expression ( $\chi^2$ =20.590,  $\rho$ <0.01). Patients with high expression of *LDHA* ( $\chi^2$ =3.856,  $\rho$ =0.049), *SLC16A1* ( $\chi^2$ =3.978,  $\rho$ =0.046) and *SLC16A3* ( $\chi^2$ =5.008,  $\chi^2$ =0.025) had lower cumulative survival rates. *SLC16A1* (*HR*=1.894, 95%*Cl*: 1.246-2.878,  $\chi^2$ =0.003) and *SLC16A3* (*HR*=1.769, 95%*Cl*:1.009-2.847,  $\chi^2$ =0.019) were the independent risk factors for overall survival (OS) in breast cancer patients. **Conclusion** *LDHA*, *SLC16A1* and *SLC16A3* are associated with certain pathologic features and poorer prognosis of breast cancer, which may provide new prognostic indicators and therapeutic targets for breast cancer treatment.

**Keywords:** Breast cancer; Glucose metabolism; Lactate dehydrogenase A; Solute carrier family 16 member 1; Solute carrier family 16 member 3; Overall survival

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The incidence and mortality of breast cancer in China are gradually increasing and are expected to continue rising [1]. According to the 2020 Global Cancer Observatory, breast cancer has become the most common cancer worldwide [2]. Tumor cells accelerate the uptake and utilization of various nutrients [3]. They tend to preferentially obtain energy through anaerobic glycolysis, a phenomenon known as the Warburg effect [4]. Although the efficiency of glycolysis in producing ATP is relatively low, the rate is much higher than that of oxidative phosphorylation [5]. Moreover, the intermediate products of glycolysis play significant roles in inhibiting cell apoptosis, promoting cell biosynthesis, and generating signaling molecules [6]. The lactate produced by anaerobic glycolysis is transported outside the cell by lactate transporters, maintaining the weak acidity of the tumor microenvironment, which is more conducive to tumor growth [7-8].

The lactate dehydrogenase A (*LDHA*) catalyzes the reduction of pyruvate to lactate, a key step in glycolysis. The solute carrier family 16 members (*SLC16A*) encode monocarboxylate transporters (MCT), mainly responsible for transporting lactate generated from intracellular

metabolism to the extracellular environment, preventing intracellular lactate accumulation and maintaining the acidic environment outside tumor cells. They can also transport extracellular lactate into cells to provide metabolic substances for tumor cells [9]. SLC16A1 and SLC16A3 encode MCT1 and MCT4, respectively. MCT1 primarily depends on the concentration of lactate and protons inside and outside the cell for lactate transport, while MCT4 mainly transports lactate produced in the glycolytic pathway out of the cells.

This study aims to analyze the expression of glucose metabolism genes in breast cancer tissues using The Cancer Genome Atlas Database (TCGA) to provide new insights into the prevention and treatment of breast cancer.

### 1 Materials and Methods

### 1.1 Source of Sample Data

Data related to breast cancer were downloaded from The Cancer Genome Atlas Database (TCGA), including two groups: the first group includes 113 samples of adjacent normal breast tissue and cancerous breast tissue,

which contain data on the expression of all gene mRNA; the second group includes 1,097 female breast cancer patients with clinical and pathological information.

The integration of the first and second groups resulted in the third group, containing clinicopathological characteristics, follow-up and death time, and mRNA expression data in 1,060 female breast cancer patients. The clinical and pathological features of data mainly include two types. One type is the features that do not need to be obtained from tumor tissue, mainly including

race, age, menopausal status, and surgical method. Another type is the features that need to be obtained from tumor tissue, mainly including tumor size (T), lymph node metastasis (N), distant metastasis (M), tumor staging, distributions of breast lesion locations, and anatomical quadrants. The data also includes overall survival (OS) and their survival status of patients, with the endpoint being patient death. The total mortality was 14.06%. [Table 1]

Tab.1 Characteristics and mortality of 1,060 patients

Clinical characteristics	Mortality (%)	Clinical characteristics	Mortality (%)
Age		M stage	
≤58 years	12.24(66/539)	M0	13.51 (119/881)
>58years	15.93(83/521)	M1	77.27 (17/22)
Race		Mx	8.28 (13/157)
White people	14.83(109/735)	Tumor stage	
Asian	51.72(30/58)	I	8.89 (16/180)
Black people or others	1.65(3/182)	п	10.67 (64/600)
Deletion	8.24(7/85)	Ш	18.57 (44/237)
Menopausal state		IV	75.00(15/20)
Premenopausal	8.04(18/224)	x	50.00(6/12)
Postmenopausal	13.17(89/676)	Deletion	36.36(4/11)
Perimenopausal	2.63(1/38)	ER state	
Deletion	33.61(41/122)	Positive	12.84(90/701)
Surgical method		Negative	17.70(37/209)
Simple mastectomy	8.63(17/197)	Deletion	14.67(22/150)
Modified radical mastectomy	18.59(58/312)	PR state	
Breast tumor resection	10.00(24/240)	Positive	13.14(80/609)
Others	14.67(38/259)	Negative	16.11(48/298)
Deletion	23.08(12/52)	Deletion	13.73(21/153)
Margin state		HER-2 state	
Positive	25.33(19/75)	Positive	12.57(22/175)
Negative	9.70(86/887)	Negative	10.63(69/649)
Unclear	19.35(6/31)	Deletion	24.58(58/236)
Deletion	56.72(38/67)	Distributions of breast lesions	
T stage		Right side	13.52(68/503)
T1	11.87(33/278)	Left side	14.54(81/557)
T2	12.32(75/609)	Tumor quadrant position	
Т3	18.80(25/133)	Right inner upper	9.43(5/53)
T4	40.54(15/37)	Right inner lower	15.38(4/26)
Tx	33.33(1/3)	Right outer upper	9.85(20/203)
N stage		Right outer lower	14.58(7/48)
N0	8.72(41/470)	Left inner upper	12.96(7/54)
N1	16.15(62/384)	Left inner lower	27.27(6/22)
N2	18.49(22/119)	Left outer upper	12.20(20/164)
N3	21.43(15/70)	Left outer lower	10.91(6/55)
Nx	52.94(9/17)	Unclear	17.01(74/435)

Note: x indicates unclear staging; ER, estrogen receptor; PR, progesterone receptor; HER-2, human epidermal growth factor receptor 2.

### 1.2 Statistical Methods

SPSS 27.0 and GraphPad Prism 9.5 were used for data processing. Data of gene expression were numerical variables, paired sample t-tests were used to compare gene expression in cancerous and adjacent normal tissues. A critical value was determined from 10% to 90% of the data at 10% intervals, with the minimum P-value corresponding to the critical value in survival analysis as the gene expression threshold.  $\chi^2$  tests were used to explore the correlation between gene expression levels and clinicopathological characteristics. Kaplan-Meier survival analysis and log-rank tests were performed to analyze differences between the groups. Cox proportional hazard models were used for univariate and multivariate analyses. P<0.05 was considered statistically significant.

### 2 Results

### 2.1 Differential Gene Expression

LDHA (430.55 $\pm$ 233.15 vs 255.79 $\pm$ 59.83, t=7.692, P<0.01), SLC16A1 (31.49 $\pm$ 35.72 vs 23.66 $\pm$ 8.97, t=2.294, P=0.024), and SLC16A3 (11.75 $\pm$ 10.30 vs 3.03 $\pm$ 2.45, t=9.481, P<0.01) were significantly upregulated in cancerous tissues compared to adjacent normal tissues in the same patient.

- 2.2 Correlation Between Gene Expression and Clinicopathological Characteristics
- 2.2.1 Correlation between *LDHA* expression and clinicopathological characteristics

*LDHA* expression levels showed significant differences in distant metastasis (M) staging, ER expression status, and HER-2 status (P<0.05), while there was no significant difference in other pathological characteristics (P>0.05). [Table 2]

2.2.2 Correlation between *SLC16A1* expression and clinicopathological characteristics

SLC16A1 expression levels showed significant differences in age, ER expression status, and PR expression status (P<0.05), while there was no

significant difference in other pathological characteristics (*P*>0.05). **[Table 2]** 

2.2.3 Correlation between *SLC16A3* expression and clinicopathological characteristics

SLC16A3 expression levels showed significant differences in ER expression status and PR expression status (P<0.05), while there was no significant difference in other pathological characteristics (P>0.05). [**Table 2**]

2.3 Correlation of ESR1 and PGR with the Above Genes

ESR1 showed a significant negative correlation with SLC16A1 (r=-0.230, P<0.01) and SLC16A3 (r=-0.143, P<0.01). Similarly, PGR exhibited a significant negative correlation with SLC16A1 (r=-0.123, P<0.01) and SLC16A3 (r=-0.110, P<0.01). No significant correlation was observed between ESR1, PGR, and LDHA expression levels. [Figure 1]

### 2.4 Survival Analysis

Kaplan-Meier survival analysis indicated that the OS of patients with high expression of *LDHA* ( $\chi^2$ =3.856, P=0.049), *SLC16A1* ( $\chi^2$ =3.978, P=0.046), and *SLC16A3* ( $\chi^2$ =5.008, P=0.025) were lower than those of patients with low expression. [Figure 2]

### 2.5 Cox Proportional Hazard Analysis

Univariable Cox regression analysis reveals that the expression levels of SLC16A1 and SLC16A3, age, tumor size (T), lymph node metastasis (N), distant metastasis (M), and clinical staging influence OS in breast cancer patients (P<0.05). Multivariable Cox analysis shows that high expression of SLC16A1, high expression of SLC16A3, advanced age, lymph node metastasis (N1-N3), distant metastasis (M1), and clinical staging III-IV were independent risk factors for OS in breast cancer patients (P<0.05). Tumor size (T3-T4) was not an independent risk factor for OS in breast cancer patients (P>0.05). [Table 3]

Tab.2 Association of LDHA, SLC16A and SLC16A3 expression with clinical characteristics in 1,060 breast cancer patients (case)

	LDHA			SLC16A1				SLC16A3				
Clinical			χ <sup>2</sup>	P			$\chi^2$	P			$\chi^2$	P value
characteristics	Low	High	value	value	Low	High	value	value	Low	High	value	r value
	expression	expression			expression	expression			expression	expression		
	(n=878)	(n=182)			(n=323)	(n=737)			(n=917)	(n=143)		

Age			0.527	0.468			8.040	0.005			0.238	0.625
≤ 58 years	266	97			143	396			469	70		
>58 years	273	85			180	341			448	73		
Menopausal state			0.010	0.92			0.250	0.617			2.235	0.135
Premenopausal	187	37			66	158			201	23		
Postmenopausal/ Perimenopausal	594	120			223	491			613	101		
Deletion	97	25			34	88			103	19		
Margin state			0.464	0.496			0.058	0.810			2.249	0.134
Positive	60	15			24	51			69	6		
Negative	737	150			272	615			761	126		
Unclear	30	1			12	19			28	3		
Deletion	51	16			15	52			59	8		
T stage			1.713	0.191			1.721	0.190			0.486	0.486
T1-T2	741	146			263	624			765	122		
T3-T4	135	35			59	111			150	20		
Deletion	2	1			1	2			2	1		
N stage			0.917	0.338			2.874	0.09			0.201	0.654
N0	384	86			129	341			404	66		
N1-N3	481	92			185	388			498	75		
Deletion	13	4			9	8			15	2		
M stage			5.560	0.018			0.101	0.751			0.049	0.826
М0	731	150			268	613			756	125		
M1	14	8			6	16			18	4		
Deletion	133	24			49	108			143	14		
Tumor stage			1.495	0.221			2.291	0.13			0.290	0.590
I - II	654	126			228	552			678	102		
III-IV	207	50			88	169			220	37		
Unclear	10	2			7	5			10	2		
Deletion	7	4			0	11			9	2		
ER state			8.532	0.003			17.428	< 0.01			22.447	< 0.001
Positive	597	104			229	472	12		624	77	,,	*****
Negative	160	49			37	172			159	50		
Deletion	121	29			57	93			134	16		
PR state	121	2)	2.904	0.088	37	75	5.486	0.019	154	10	20.59	< 0.001
Positive	514	95	2.504	0.000	193	416	3.400	0.017	546	63	20.37	·0.001
Negative	238	60			72	226			234	64		
Deletion	126	27			58	95			137	17		
HER-2 state	120	21	4.418	0.036	36	)3	2.871	0.090	137	17	0.592	0.442
Positive	136	39	4.410	0.030	59	116	2.6/1	0.090	147	20	0.392	0.442
Negative	548				39 179	116 479			560	28		
Deletion		101								89		
Distributions of	194	42			85	142			210	26		
breast lesions Right side	416	87	0.011	0.917	149	354	0.326	0.568	445	58	3.15	0.076
Left side	462	95			174	383			472	85		
Tumor quadrant position	702	93	0.844	0.839	1/4	303	0.846	0.839	7/2	63	4.564	0.207
Inner upper	91	16			31	76			96	11		

Inner lower	38	10	17	31	37	11
Outer upper	304	63	107	260	309	58
Outer lower	86	17	30	73	89	14
Deletion	359	76	138	297	386	49

Fig. 1 Heat map of gene correlation coefficients

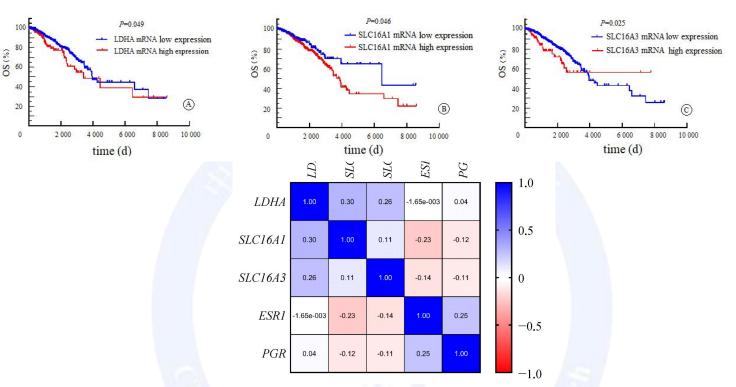


Fig.2 Kaplan Meier survival analysis of breast cancer patients with different gene mRNA expression levels

Tab.3 Univariable and multivariable Cox proportional hazard model of survival in 1,060 breast cancer patients

	Univariable Cox re	Multivariable Cox regression		
Clinical characteristics	HR (95%CI)	P value	Multivariable Cox r HR (95%CI)  1.894(1.246-2.878) 1.769(1.099-2.847) 2.014(1.398-2.903) 1.016(0.612-1.687) 1.616(1.033-2.527) 3.121(1.632-5.969) 1.718(1.000-2.953)	P value
High expression of SLC16A1	1.457(1.004-2.114)	0.047	1.894(1.246-2.878)	0.003
High expression of SLC16A3	1.640(1.059-2.542)	0.027	1.769(1.099-2.847)	0.019
Age>58 years	1.782(1.288-2.467)	< 0.001	2.014(1.398-2.903)	< 0.001
T3-T4 stage	1.769(1.227-2.550)	0.002	1.016(0.612-1.687)	0.950
N1-N3 stage	2.170(1.507-3.126)	< 0.001	1.616(1.033-2.527)	0.036
Distant metastasis M1	2.445(1.482-4.066)	< 0.001	3.121(1.632-5.969)	< 0.001
Clinical staging III-IV	2.659(1.899-3.724)	< 0.001	1.718(1.000-2.953)	$0.050^{a}$

Note: <sup>a</sup> P=0.049,97<0.050 when the number is accurate to 5 decimal places, at the threshold of significance.

#### 3 Discussion

Research suggested that high expression of *LDHA* in tumor tissues was associated with adverse prognosis. Knockout or inhibition of *LDHA* could impede tumor cell growth [12-13]. Zhao *et al.* [14] found a significant association between high expression of *LDHA* and high histological grade, lymph node metastasis, tumor staging of breast cancer. Negative ER expression showed higher

LDHA expression. Fantin et al. [12] showed that inhibiting LDHA reduced the transformation of malignant tumors and delayed tumor formation. Our study confirms an association between high LDHA mRNA expression / distant metastasis and ER and HER-2 expression. In addition, high LDHA mRNA expression was associated with worse OS in patients.

Studies have shown a positive association between high expression of MCT1 and patient prognosis in non-

small cell lung cancer, head-and-neck cancer, and other cancers. The OS rate of the high SLC16A1 expression group was significantly higher than that of the low expression group [15-16]. On the contrary, some scholars found that the high expression of SLC16A1 was related to the adverse clinical results of urinary system tumors, primary neuroblastoma and breast cancer [17-18]. Johnson et al. [19] found a higher expression of MCT1 in triple negative breast cancer compared to other subtypes, and overexpression of MCT1 was associated with an increased risk of tumor growth and recurrence. In this study, breast cancer patients in the high SLC16A1 expression group had worse OS. The difference in the above results may be due to the fact that lactate transport by MCT1 depends on the pH inside and outside the cells [10]. There are specific differences in the pH gradient inside and outside the cells of different tumors. The sample size is insufficient, which makes the statistical analysis inevitably deviate, and further sample collection may be required for verification. Our study suggests that SLC16A1 is negatively correlated with ESR1 and PGR, and higher SLC16A1 may predict low expression of ER and PR, so that patients lose effective endocrine therapeutic targets, resulting in worse OS.

Khan *et al.* [20] treated neuroblastoma with MCT1 inhibitor, which disrupted lactate homeostasis and NADH/NAD<sup>+</sup> ratio of cells, inhibited the growth of cancer cells, and highly synergized with *LDHA* inhibitor to reduce cell viability. Hou *et al.* [21] found that inhibiting MCT1 could overcome the resistance of breast cancer cells to Paclitaxel. The above results showed the potential of targeting the corresponding proteins of *LDHA* and *SLC16A1* to improve the prognosis of patients.

MCT4 is highly expressed in prostate cancer, intrahepatic cholangiocarcinoma, lung cancer, bladder cancer and liver cancer, and is closely related to the proliferation, invasion and metastasis of tumor cells. Its high expression is associated with worse patient OS [22-26]. This study confirmed it. Silencing of MCT4 in prostate tumors significantly reduced cell proliferation, migration, and invasion [22]. Studies found an upregulated expression of MCT4 in breast cancer, and the higher MCT4 expression was associated with poor prognosis, immune cell infiltration and glycolysis rate-limiting enzyme [18]. *SLC16A3* is expected to become a potential prognostic indicator and target for tumor therapy to assist the treatment of breast tumors.

The results of this study showed that the expression levels of *LDHA*, *SLC16A1* and *SLC16A3* were higher in the ER and PR negative group. Zhao *et al.* [14] also found an increased expression of *LDHA* in the ERnegative group. *ESR1* and *PGR* were significantly negatively correlated with *SLC16A1* and *SLC16A3*. The relationship between the expression of ER, PR and the above genes needs to be further studied.

The study has some limitations. Firstly, the dataset has only a small proportion of Asian patients, which may be biased by gene expression in our country. Secondly, although TCGA database has accurate sequencing, good quality and multi-omics data [27-29], the 1,060 breast cancer samples obtained from TCGA database in this study still have insufficient data, and clinical samples can be collected later to expand the sample further. Finally, the study solely focuses on mRNA levels, and future research could benefit from incorporating other omics levels for a more comprehensive analysis. Despite these limitations, the study draws relevant conclusions.

In conclusion, *LDHA*, *SLC16A1* and *SLC16A3* genes are highly expressed in breast cancer tissues and negatively correlated with OS of breast cancer patients. *SLC16A1* and *SLC16A3* are independent risk factors for OS in breast cancer patients. These genes are expected to become prognostic indicators of breast cancer, which may provide a new target for the subsequent treatment of breast cancer.

#### Conflict of interest: None

#### Reference

- Lei SY, Zheng RS, Zhang SW, et al. Breast cancer incidence and mortality in women in China: temporal trends and projections to 2030[J]. Cancer Biol Med, 2021, 18(3): 900-909.
- [2] Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries[J]. CA Cancer J Clin, 2018, 68(6): 394-424.
- [3] Sousa B, Pereira J, Paredes J. The crosstalk between cell adhesion and cancer metabolism[J]. Int J Mol Sci, 2019, 20(8): 1933.
- [4] Lebelo MT, Joubert AM, Visagie MH. Warburg effect and its role in tumourigenesis[J]. Arch Pharm Res, 2019, 42(10): 833-847.
- [5] Locasale JW, Cantley LC. Altered metabolism in cancer[J]. BMC Biol, 2010, 8: 88.
- [6] Hamanaka RB, Chandel NS. Targeting glucose metabolism for cancer therapy[J]. J Exp Med, 2012, 209(2): 211-215.
- [7] Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism[J]. Cell Metab, 2016, 23(1): 27-47.
- [8] Tu VY, Ayari A, O'Connor RS. Beyond the lactate paradox: how lactate and acidity impact T cell therapies against cancer[J]. Antibodies, 2021, 10(3): 25
- [9] Pereira-Nunes A, Afonso J, Granja S, et al. Lactate and lactate transporters as key players in the maintenance of the Warburg effect[J]. Adv Exp Med Biol, 2020, 1219: 51-74.
- [10] Pérez-Escuredo J, Van Hée VF, Sboarina M, et al. Monocarboxylate transporters in the brain and in cancer[J]. Biochim Biophys Acta, 2016, 1863(10): 2481-2497.
- [11] Benjamin D, Robay D, Hindupur SK, et al. Dual Inhibition of the Lactate Transporters MCT1 and MCT4 Is Synthetic Lethal with Metformin due to NAD+ Depletion in Cancer Cells[J]. Cell Rep, 2018, 25(11): 3047-3058.e4.
- [12] Fantin VR, St-Pierre J, Leder P. Attenuation of LDH-a expression uncovers a link between glycolysis, mitochondrial physiology, and tumor maintenance[J]. Cancer Cell, 2006, 9(6): 425-434.
- [13] Vander Heiden MG. Targeting cancer metabolism: a therapeutic window opens[J]. Nat Rev Drug Discov, 2011, 10(9): 671-684.
- [14] Zhao XK, Li W, Liu W, et al. Effect of LDHA on the evolution of breast cancer and SUMOylation modification[J]. Chin J Clin Exp Pathol, 2022, 38(4): 427-431.
- [15] Eilertsen M, Andersen S, Al-Saad S, et al. Monocarboxylate transporters 1-4 in NSCLC: MCT1 is an independent prognostic marker for survival[J]. PLoS One, 2014, 9(9): e105038.
- [16] Zhang Q, Tang Z. SLC16A1 gene expression and prognosis in oral cancer and other head and neck cancer: an analysis based on oncomine database[J]. J Nanchang Univ Med Sci, 2018, 58(4): 10-14, 43.
- [17] Zhang L, Song ZS, Wang ZS, et al. High expression of SLC16A1 as a



- biomarker to predict poor prognosis of urological cancers[J]. Front Oncol, 2021, 11: 706883.
- [18] Yuan C, Zhang J, Lou JJ, et al. Comprehensive Analysis of Monocarboxylate Transporter 4 (MCT4) expression in breast cancer prognosis and immune infiltration via integrated bioinformatics analysis[J]. Bioengineered, 2021, 12(1): 3850-3863.
- [19] Johnson JM, Cotzia P, Fratamico R, et al. MCT1 in invasive ductal carcinoma: monocarboxylate metabolism and aggressive breast cancer[J]. Front Cell Dev Biol, 2017, 5: 27.
- [20] Khan A, Valli E, Lam H, et al. Targeting metabolic activity in high-risk neuroblastoma through Monocarboxylate Transporter 1 (MCT1) inhibition[J]. Oncogene, 2020, 39(17): 3555-3570.
- [21] Hou L, Zhao Y, Song GQ, et al. Interfering cellular lactate homeostasis overcomes Taxol resistance of breast cancer cells through the microRNA-124-mediated lactate transporter (MCT1) inhibition[J]. Cancer Cell Int, 2019, 19: 193.
- [22] Sun Q, Hu LL, Fu Q. MCT4 promotes cell proliferation and invasion of castration-resistant prostate cancer PC-3 cell line[J]. EXCLI J, 2019, 18: 187-194.
- [23] Dong LQ, Lu DY, Chen R, et al. Proteogenomic characterization identifies clinically relevant subgroups of intrahepatic cholangiocarcinoma[J]. Cancer Cell, 2022, 40(1): 70-87.e15.

- [24] Xue L, Liu JY, Xie JH, et al. Prognostic value of SLC16A3(MCT4) in lung adenocarcinoma and its clinical significance[J]. Int J Gen Med, 2021, 14: 8413-8425.
- [25] Zhao Y, Zhao B, Yan WH, et al. Integrative analysis identified MCT4 as an independent prognostic factor for bladder cancer[J]. Front Oncol, 2021, 11: 704857.
- [26] Mukai Y, Yamaguchi A, Sakuma T, et al. Involvement of SLC16A1/MCT1 and SLC16A3/MCT4 in 1-lactate transport in the hepatocellular carcinoma cell line[J]. Biopharm Drug Dispos, 2022, 43(5): 183-191.
- [27] Tomczak K, Czerwińska P, Wiznerowicz M. The Cancer Genome Atlas (TCGA): an immeasurable source of knowledge[J]. Contemp Oncol, 2015, 19(1A): A68-A77.
- [28] Zhang Z, Li H, Jiang S, et al. A survey and evaluation of Web-based tools/databases for variant analysis of TCGA data[J]. Brief Bioinform, 2019, 20(4): 1524-1541.
- [29] Zhang M, Zhou LZ, Sun Z, et al. Analysis of the expression and clinical significance of POSTN in head and neck squamous carcinoma based on TCGA database[J]. Chin J Gen Pract, 2022, 20(6): 1066-1070.

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· 论 著·

# 基于 TCGA 数据库分析葡萄糖代谢相关基因与 乳腺癌临床病理特性及预后相关性

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摘要:目的 探究乳酸脱氢酶 A(LDHA)、溶质载体家族 16 成员(SLC16A) 1 及 SLC16A3 基因与乳腺癌病理特征 及预后的关系。方法 获取肿瘤基因组图谱数据库(TCGA) 中 1 060 位乳腺癌患者的组织样本,分析 LDHA、SLC16A1 及 SLC16A3 基因表达量与乳腺癌临床病理特征及预后的关联性,用 Kaplan-Meier 生存分析绘制生存曲线,Cox 比例风险回归模型进行单因素和多因素生存预后分析。结果 LDHA 表达与远处转移(M 分期)( $X^2 = 5.560$ , P = 0.018)、雌激素受体(ER)表达( $X^2 = 8.532$ , P = 0.003)、人表皮生长因子受体-2(ER-2) 表达( $X^2 = 4.418$ , P = 0.036)相关;SLC16A1 表达与年龄( $X^2 = 8.040$ , Y = 0.005)、ER 表达( $Y^2 = 17.428$ , Y = 0.01)、Y = 0.010,有法。Y = 0.010,相关;Y = 0.010,相关;Y = 0.010,相关;Y = 0.010,是不是这(Y = 0.010,是一个。Y = 0.011,是一个。Y = 0.012,是一个。Y = 0.013,是一个。Y = 0.013,是一个。Y = 0.014,是一个。Y = 0.014,是一个。Y = 0.015,是一个。Y = 0.015,是一个。

### Glucose metabolism-related genes with clinicopathological characteristics and prognosis of breast cancer: an analysis based on TCGA database

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**Abstract: Objective** To investigate the relationship between the expression of *LDHA*, *SLC16A1* and *SLC16A3* genes and pathologic features and prognosis in breast cancer. **Methods** Tissue samples from 1 060 breast cancer patients in The Cancer Genome Atlas (TCGA) were obtained. The association of *LDHA*, *SLC16A1* and *SLC16A3* gene expressions with clinicopathological features and prognosis of breast cancer were analyzed. Survival curve were drawn by Kaplan-Meier survival analysis, and univariable and multivariable survival prognosis were analyzed by Cox proportional hazard regression model. **Results** *LDHA* expression was associated with distant metastasis (M stage) ( $\chi^2 = 5.560$ , P = 0.018), estrogen receptor (ER) expression ( $\chi^2 = 8.532$ , P = 0.003), and human epidermal growth factor receptor 2 (HER-2) expression ( $\chi^2 = 4.418$ , P = 0.036); *SLC16A1* expression correlated with age ( $\chi^2 = 8.040$ , P = 0.005), ER expression ( $\chi^2 = 17.428$ , P < 0.01), and progesterone receptor (PR) expression ( $\chi^2 = 5.486$ ,  $\chi^2 = 0.019$ ). *SLC16A3* expression correlated with ER expression ( $\chi^2 = 22.447$ ,  $\chi^2 = 0.019$ ), PR expression ( $\chi^2 = 20.590$ ,  $\chi^2 = 0.019$ ). Patients with high expression of *LDHA* ( $\chi^2 = 3.856$ ,  $\chi^2 = 0.049$ ), *SLC16A1* ( $\chi^2 = 3.978$ ,  $\chi^2 = 0.046$ ) and *SLC16A3* ( $\chi^2 = 5.008$ ,  $\chi^2 = 0.025$ ) had lower cumulative survival rates. *SLC16A1* ( $\chi^2 = 1.894$ ,  $\chi^2 = 0.046$ ) and *SLC16A3* ( $\chi^2 = 0.003$ ) and *SLC16A3* ( $\chi^2 = 0.005$ ) had lower cumulative survival rates. *SLC16A1* ( $\chi^2 = 0.005$ ) were the independent risk factors for overall survival in breast cancer patients.

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QR code for English version

**Conclusion** *LDHA*, *SLC16A1* and *SLC16A3* are associated with certain pathologic features and poorer prognosis of breast cancer, which may provide new prognostic indicators and therapeutic targets for breast cancer treatment.

**Keywords**: Breast cancer; Glucose metabolism; Lactate dehydrogenase A; Solute carrier family 16 member 1; Solute carrier family 16 member 3; Overall survival

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中国乳腺癌的发病率与死亡率逐步上升,且预计呈现持续上升趋势<sup>[1]</sup>。根据 2020 年全球各类癌症登记数据显示,乳腺癌已成为全球发病率最高的肿瘤<sup>[2]</sup>。肿瘤细胞会加快对各类营养物质的摄取与转化利用<sup>[3]</sup>,肿瘤细胞偏向于通过无氧酵解途径获取能量,这个现象被称为瓦博格(Warburg)效应<sup>[4]</sup>。虽然糖酵解生成三磷酸腺苷的效率较低,但速率却远远高于氧化磷酸化<sup>[5]</sup>;且糖酵解的中间产物在抑制细胞凋亡、促进细胞生物合成、产生信号分子等许多方面发挥了巨大作用<sup>[6]</sup>,无氧酵解所产生的乳酸经过乳酸转运蛋白转运至胞外,维持肿瘤微环境的弱酸性,更适合肿瘤的生长<sup>[7-8]</sup>。

乳酸脱氢酶 A(lactate dehydrogenase A, LDHA) 基因编码 LDHA,它催化丙酮酸还原成乳酸,这是糖酵解的关键步骤。溶质载体家族 16 成员(solute carrier family 16 member, SLC16A)基因编码单羧酸转运蛋白(monocarboxylate transporter, MCT)。MCT 主要是将胞内代谢产生的乳酸转运至胞外,维持肿瘤细胞外的酸性环境;它还可将胞外的乳酸转运至胞内为肿瘤细胞提供代谢物质<sup>[9]</sup>。SLC16A1 和 SLC16A3 分别编码 MCT1 和 MCT4, MCT1(SLC16A1 基因编码)对于乳酸的输入和输出主要取决于细胞内外的乳酸和质子浓度<sup>[10]</sup>;MCT4(SLC16A3 基因编码)主要作用是将糖代谢途径中产生的乳酸转运至细胞外<sup>[11]</sup>。本文用 TCGA 数据库分析乳腺癌组织中葡萄糖代谢基因的表达,为乳腺癌的预防及治疗提供新论点及思路。

### 1 材料与方法

1.1 标本数据来源 下载肿瘤基因组图谱数据库(the Cancer Genome Atlas Database, TCGA)中乳腺癌相关数据。数据为以下 2 组:第 1 组数据包括 113 例患者癌旁乳腺组织及其癌变乳腺组织样本,这些样本包含了所有基因 mRNA 表达的数据;第 2 组数据包括临床病理信息数据的 1 097 名女性乳腺癌患者资料。

第1组数据与第2组数据整合出第3组数据:包含临床病理特征、随访及死亡时间和基因 mRNA 表达数据的1060名女性乳腺癌患者资料。数据的临床病理特征主要包括两类,一类是无需从肿瘤组织中

获取的特征,主要包括:种族、年龄、绝经状态、手术方式;另一类是需要从肿瘤组织获取的特征,主要包括:肿瘤大小(T)、淋巴结转移(N)、远处转移(M)、肿瘤分期、肿瘤左右位置以及解剖象限。数据还包括患者总生存时间(overall survival, OS)和患者的生存状态,终点为患者死亡。总死亡率为14.06%。见表1。

表 1 1060 例患者的临床病理特征及其死亡率 **Tab. 1** Characteristics and death rates of 1060 patients

临床特征	死亡率(%)	临床特征	死亡率(%)
年龄		M 分期	
≤58 岁	12.24(66/539)	MO	13.51(119/881)
>58 岁	15.93(83/521)	M1	77.27(17/22)
种族		Mx	8.28(13/157)
白人	14.83(109/735)	肿瘤分期	
亚洲人	51.72(30/58)	I	8.89(16/180)
黑人或其他	1.65(3/182)	II	10.67(64/600)
缺失	8.24(7/85)	Ⅲ	18.57(44/237)
绝经状态		IV	75.00(15/20)
绝经前	8.04(18/224)	X	50.00(6/12)
绝经后	13.17(89/676)	缺失	36.36(4/11)
围绝经期	2.63(1/38)	ER 状态	
缺失	33.61(41/122)	阳性	12.84(90/701)
手术方式		阴性	17.70(37/209)
单纯式乳房切除术	8.63(17/197)	缺失	14.67(22/150)
改良乳腺根治术	18.59(58/312)	PR 状态	
乳腺肿瘤切除术	10.00(24/240)	阳性	13.14(80/609)
其他	14.67(38/259)	阴性	16.11(48/298)
缺失	23.08(12/52)	缺失	13.73(21/153)
切缘状态		HER-2 状态	
阳性	25.33(19/75)	阳性	12.57(22/175)
阴性	9.70(86/887)	阴性	10.63(69/649)
不明确	19.35(6/31)	缺失	24.58(58/236)
缺失	56.72(38/67)	肿瘤位置	
T 分期		右侧	13.52(68/503)
T1	11.87(33/278)	左侧	14.54(81/557)
T2	12.32(75/609)	肿瘤象限位置	
T3	18.80(25/133)	右内上象限	9.43(5/53)
T4	40.54(15/37)	右内下象限	15.38(4/26)
Tx	33.33(1/3)	右外上象限	9.85(20/203)
N 分期		右外下象限	14.58(7/48)
NO	8.72(41/470)	左内上象限	12.96(7/54)
N1	16.15(62/384)	左内下象限	27.27(6/22)
N2	18.49(22/119)	左外上象限	12.20(20/164)
N3	21.43 (15/70)	左外下象限	10.91 (6/55)
Nx	52.94(9/17)	未明确	17.01(74/435)

注:分期中 x 表示未知; ER 为雌激素受体; PR 为孕激素受体; HER-2 为人表皮生长因子受体。

1.2 统计学方法 使用 SPSS 27.0、Graphpad Pism 9.5 对数据进行处理。基因表达数据属于数值变量,使用成对样本 t 检验比较癌变组织及癌旁组织中的基因表达。在数据的 10%~90%之间每隔 10 个数取一个临界值,以生存分析结果中得出最小 P 值对应的临界值作为基因的表达阈值将基因表达分为高表达组和低表达组。用 $\mathcal{X}^2$  检验分析基因的表达水平与临床病理特征的关联性。对不同组别进行 Kaplan-Meier 生存分析,绘制生存曲

线,运用 log-rank 检验分析其差异性。Cox 比例风险回归模型进行单因素与多因素分析,将差异有统计学意义的单因素进行多因素分析。P<0.05 为差异具有统计学意义。

### 2 结 果

2.1 基因的差异表达 同一患者的癌变组织 *LDHA* (430.55±233.15 vs 255.79±59.83, t = 7.692, P < 0.01)、SLC16A1 (31.49±35.72 vs 23.66±8.97, t =

2.294, P = 0.024)、SLC16A3 (11.75±10.30 vs 3.03±2.45, t = 9.481, P < 0.01) 表达均高于自身的癌旁组织,差异有统计学意义。

2.2 基因表达与乳腺癌患者临床病理特征关联性 2.2.1 *LDHA* 的表达与临床病理特征的关联性 *LDHA* 表达量的高低在远处转移(M分期)、ER 表达状态、HER-2 表达状态中差异有统计学意义(P<0.05),在其他病理特征中差异无统计学意义(P>0.05)。见表 2。

表 2 1060 名乳腺癌患者 LDHA、SLC16A 及 SLC16A3 表达与临床特征的关联性 (例) **Tab. 2** Association of LDHA, SLC16A and SLC16A3 expression with clinical characteristics in 1060 breast cancer patients (case)

	LDHA				16A1			SLC16A3				
病理特征	低表达 (n=878)	高表达 (n=182)	X <sup>2</sup> 值	P 值	低表达 (n=323)	高表达 (n=737)	X <sup>2</sup> 值	P 值	低表达 (n=917)	高表达 (n=143)	X <sup>2</sup> 值	P 值
年龄												
≤58 岁	442	97	0.527	0.468	143	396	8.04	0.005	469	70	0.238	0.625
>58 岁 绝经状态	436	85	0.327	0.408	180	341	8.04	0.003	448	73	0.238	0.02.
绝经前	187	37			66	158			201	23		
绝经后或围绝经	594	120	0.010	0.920	223	491	0.250	0.617	613	101	2.235	0.13
缺失 切缘状态	97	25			34	88			103	19		
阳性	60	15			24	51			69	6		
阴性	737	150	0.464	0.496	272	615	0.058	0.810	761	126	2.249	0.13
不明确	30	1	0.404	0.470	12	19	0.056	0.010	28	3		
缺失 T 分期	51	16			15	52			59	8		
T1~T2	741	146			263	624			765	122		
T3~T4	135	35	1.713	0.191	59	111	1.721	0.190	150	20	0.486	0.486
缺失 N 分期	2	1			1	2			2	1		
NO	384	86			129	341			404	66		
N1 ~ N3	481	92	0.917	0.338	185	388	2.874	0.090	498	75	0.201	0.654
缺失 M 分期	13	4			9	8			15	2		
MO	731	150			268	613			756	125		
M1	14	8	5.560	0.018	6	16	0.101	0.751	18	4	0.049	0.826
缺失 肿瘤分期	133	24			49	108			143	14		
I ~ II	654	126			228	552			678	102		
<b>Ⅲ</b> ~ <b>Ⅳ</b>	207	50	1.495	0.221	88	169	2.291	0.130	220	37	0.290	0.59
不明确	10	2	1.473	0.221	7	5	2.291	0.130	10	2	0.290	0.55
缺失	7	4			0	11			9	2		
ER												
阳性	597	104			229	472	.= .=.		624	77		
阴性	160	49	8.532	0.003	37	172	17.428	< 0.001	159	50	22.447	< 0.00
缺失 PR	121	29			57	93			134	16		
阳性	514	95			193	416			546	63		
阴性	238	60	2.904	0.088	72	226	5.486	0.019	234	64	20.590	< 0.00
缺失 HER-2	126	27			58	95			137	16		
阳性	136	39			59	116			147	28		
阴性	548	101	4.418	0.036	179	479	2.871	0.090	560	89	0.592	0.442
缺失 肿瘤位置	194	42			85	142			210	26		
右侧乳腺	416	87	0.011	0.017	149	354	0.226	0.560	445	58	2.150	0.07
左侧乳腺	462	95	0.011	0.917	174	383	0.326	0.568	472	85	3.150	0.076
肿瘤象限分布												
内上	91	16			31	76			96	11		
内下	38	10			17	31			37	11		
外上	304	63	0.844	0.839	107	260	0.846	0.839	309	58	4.564	0.20
外下	86	17			30	73			89	14		
缺失	359	76			138	297			386	49		

注:缺失不纳入统计分析。

- 2.2.2 *SLC16A1* 的表达与临床病理特征的关联性 *SLC16A1* 表达量的高低在年龄、ER 表达状态、PR 表达 状态中差异有统计学意义(P<0.05),在其他病理特征中差异无统计学意义(P>0.05)。见表 2。
- 2.2.3 SLC16A3 的表达与临床病理特征的关联性 SLC16A3 的表达量在 ER 表达状态、PR 表达状态中 差异有统计学意义(P<0.05),其他病理特征其表达 量高低差异无统计学意义(P>0.05)。见表 2。
- 2.3 ESRI(编码 ER 的基因)和 PGR(编码 PR 的基因)与上述基因的相关性 ESRI与 SLCI6AI(r=-0.230, P<0.01)、<math>SLCI6A3(r=-0.143, P<0.01)呈显著负相关;PGR与 SLCI6AI(r=-0.123, P<0.01)、<math>SLCI6A3(r=-0.110, P<0.01)呈现显著负相关。未见到 ESRI、PGR与 LDHA表达量之间具有显著的相关性。见图 1。

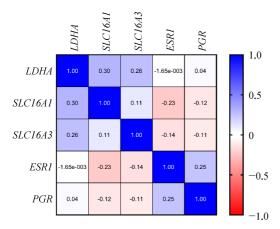
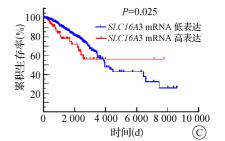


图 1 基因相关系数热图

Fig. 1 Heat map of gene correlation coefficients

P=0.046100 P=0.049100 SLC16A1 mRNA 低表达 累积生存率(%) LDHA mRNA 低表达 累积生存率(%) SLC16A1 mRNA 高表达 80 LDHA mRNA 高表达 60 60 40 20 20 2 000 4 000 6 000 8 000 10 000 2 000 4 000 6 000 8 000 10 000 0 0 时间(d) (A) 时间(d) (B)



注: A 为不同 LDHA mRNA 表达水平的 OS, B 为不同 SLC16A1 mRNA 表达水平的 OS, C 为不同 SLC16A3 mRNA 表达水平的 OS。

图 2 不同基因 mRNA 表达水平乳腺癌患者的 Kaplan-Meier 生存分析

Fig. 2 Kaplan Meier survival analysis of breast cancer patients with different gene mRNA expression levels

#### 3 讨论

有研究表明 *LDHA* 在肿瘤组织中高表达,并与肿瘤的不良的预后有关。*LDHA* 敲除<sup>[12-13]</sup> 和 *LDHA* 抑制都会抑制体外肿瘤细胞的生长<sup>[12]</sup>。赵雪可等<sup>[14]</sup>发现 *LDHA* 高表达与乳腺癌组织学高分级、淋巴结转移和肿瘤分期等相关;且 ER 阴性表现出更高的 *LDHA* 表达。

2.4 生存分析 Kaplan-Meier 生存分析结果显示  $LDHA(X^2=3.856, P=0.049)$ 、 $SLC16A1(X^2=3.978, P=0.046)$ 、 $SLC16A3(X^2=5.008, P=0.025)$ 高表达组的累积生存率较低表达组更低。见图 2。

2.5 Cox 风险回归分析 单因素 Cox 回归分析显示, SLC16A1 表达量、SLC16A3 表达量、年龄、肿瘤大小(T)、淋巴结转移(N)、远处转移(M) 及临床分期等影响患者的 OS。

对与预后有关的单因素进行多因素 Cox 分析示,SLC16A1 高表达、SLC16A3 高表达、高龄、淋巴结转移  $(N1\sim N3)$ 、远处转移 (M1) 和临床分期  $(N1\sim N3)$  和  $(N1\sim N3)$  和 (

### 表 3 1060 例乳腺癌患者生存预后影响的 Cox 单因素及多因素分析

**Tab. 3** Univariable and multivariable Cox proportional hazard regression model of survival in 1 060 breast cancer patients

病理特征	单因素 Cox 回	月	多因素 Cox 回归		
<b>州理付</b> 征	HR(95%CI)	P 值	HR(95%CI)	P 值	
SLC16A1 高表达	1.457(1.004~2.114)	0.047	1.894(1.246~2.878)	0.003	
SLC16A3 高表达	1.640(1.059~2.542)	0.027	$1.769(1.099 \sim 2.847)$	0.019	
年龄>58 岁	1.782(1.288~2.467)	< 0.001	2.014(1.398~2.903)	< 0.001	
T3~T4 分期	1.769(1.227~2.550)	0.002	$1.016(0.612 \sim 1.687)$	0.950	
N1~N3 分期	2.170(1.507~3.126)	< 0.001	1.616(1.033~2.527)	0.036	
远处转移 M1	2.445(1.482~4.066)	< 0.001	3.121(1.632~5.969)	< 0.001	
临床分期Ⅲ~Ⅳ	2.659(1.899~3.724)	< 0.001	1.718(1.000~2.953)	$0.050^{\mathrm{a}}$	

注:a 表示保留小数点后 5 位时 P=0.049 97<0.050,处于显著性临界值。

Fantin 等<sup>[12]</sup>表明,抑制 *LDHA* 可减少恶性肿瘤的转化、延迟肿瘤的形成。本研究见到 *LDHA* mRNA 高表达与远处转移和 ER、HER2 表达存在关联性,并且 *LDHA* mRNA 高表达与患者更差的 OS 相关。

有研究表明在非小细胞肺癌、头颈部癌等癌症中 MCT1 高表达与患者预后呈现正相关,*SLC16A1* 高表 达组的总体生存率明显高于低表达组<sup>[15-16]</sup>。相反, 有学者发现 SLC16A1 高表达与泌尿系统肿瘤、原发性神经母细胞瘤及乳腺癌的不良临床结果相关<sup>[17-18]</sup>。Johnson等<sup>[19]</sup>发现三阴性乳腺癌中MCT1表达高于其他亚型,且MCT1过度表达与肿瘤生长风险增加与复发有关。在本研究中,SLC16A1高表达组的乳腺癌患者有着更差的OS。上述结果的不同可能是由于MCT1转运乳酸取决于细胞内外的pH值<sup>[10]</sup>,不同肿瘤细胞内外pH值梯度存在一定差异,或是样本量不足,使统计分析结果出现不可避免的偏差,可能需要进一步收集样本验证。就本研究而言,可能是SLC16A1与ESR1、PGR呈现负相关,更高的SLC16A1可能预示着ER、PR低表达,使患者丧失有效的内分泌治疗靶点,导致患者更差的OS。

Khan 等 $^{[20]}$ 用 MCT1 抑制剂处理神经母细胞瘤,破坏细胞的乳酸稳态、还原和氧化烟酰胺腺嘌呤二核苷酸(reduced and oxidized nicotinamide adenine dinucleotide,NADH/NAD $^+$ )比率,抑制了癌细胞的生长,并且与 *LDHA* 抑制剂高度协同降低细胞活力。Hou 等 $^{[21]}$ 发现抑制 MCT1 可以克服乳腺癌细胞对紫杉醇的耐药性;上述成果显示了靶向 *LDHA* 和 *SLC16A1* 相应蛋白进而改善患者预后的潜力。

MCT4 在前列腺癌、肝内胆管癌、肺癌、膀胱癌、肝癌中高表达,且与肿瘤细胞的增殖、侵袭及转移等紧密相关,其高表达与患者更差的 OS 相关<sup>[22-26]</sup>,本研究结果与其一致。Sun 等<sup>[22]</sup>证实沉默 MCT4 可以使前列腺癌细胞的增殖、迁移和侵袭能力显著降低。有研究发现 MCT4 在乳腺癌中表达上调,并且与不良预后有关,且乳腺癌中 MCT4 的表达与免疫细胞浸润和糖酵解限速酶有关<sup>[18]</sup>,所以 SLC16A3 有望成为肿瘤治疗的一个潜在的预后指标及靶点辅助乳腺肿瘤的治疗。

本研究结果示在 ER、PR 均为阴性组中,LDHA、SLC16A1 和 SLC16A3 的表达量更高,赵雪可等<sup>[14]</sup>研究也表明 ER 阴性组的 LDHA 表达量更高,本研究结果与其一致。且 ESR1 和 PGR 与 SLC16A1 和 SLC16A3 呈现显著负相关;ER、PR 的表达与上述基因的关系有待进一步研究。

本研究存在一定的不足,首先数据中亚洲患者只占小部分,可能与我国基因表达存在偏差;其次虽然TCGA数据库具有精确的测序,良好质量的数据以及丰富的组学[27-29],但本研究从TCGA数据库获取的1060例乳腺癌样本仍存在一定的数据量不足,后续可收集临床样本进一步扩大样本量;最后,本研究仅在基因mRNA层面进行论证,未结合其他组学层面

进行研究。虽然本研究存在一定的局限性,但所得出的相关结论可为后续研究提供一定的方向。

综上所述,LDHA、SLC16A1 和 SLC16A3 基因在乳腺癌组织中高表达,与乳腺癌患者的 OS 呈负相关; SLC16A1 和 SLC16A3 是乳腺癌患者 OS 的独立危险 因素;上述基因有望成为乳腺癌的预后指标,可能为后续乳腺癌的治疗提供新的靶点。

### 利益冲突 无

### 参考文献

- [1] Lei SY, Zheng RS, Zhang SW, et al. Breast cancer incidence and mortality in women in China: temporal trends and projections to 2030 [J]. Cancer Biol Med, 2021, 18(3): 900-909.
- [2] Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries [J]. CA Cancer J Clin, 2018, 68 (6) · 394-424.
- [3] Sousa B, Pereira J, Paredes J. The crosstalk between cell adhesion and cancer metabolism[J]. Int J Mol Sci, 2019, 20(8): 1933.
- [4] Lebelo MT, Joubert AM, Visagie MH. Warburg effect and its role in tumourigenesis [J]. Arch Pharm Res, 2019, 42(10): 833-847.
- [5] Locasale JW, Cantley LC. Altered metabolism in cancer [J]. BMC Biol, 2010, 8: 88.
- [6] Hamanaka RB, Chandel NS. Targeting glucose metabolism for cancer therapy[J]. J Exp Med, 2012, 209(2): 211-215.
- [7] Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism[J]. Cell Metab, 2016, 23(1): 27-47.
- [8] Tu VY, Ayari A, O'Connor RS. Beyond the lactate paradox: how lactate and acidity impact T cell therapies against cancer [J]. Antibodies, 2021, 10(3): 25.
- [9] Pereira-Nunes A, Afonso J, Granja S, et al. Lactate and lactate transporters as key players in the maintenance of the Warburg effect [J]. Adv Exp Med Biol, 2020, 1219; 51-74.
- [10] Pérez-Escuredo J, Van Hée VF, Sboarina M, et al. Monocarboxylate transporters in the brain and in cancer [J]. Biochim Biophys Acta, 2016, 1863(10): 2481-2497.
- [11] Benjamin D, Robay D, Hindupur SK, et al. Dual inhibition of the lactate transporters MCT1 and MCT4 is synthetic lethal with metformin due to NAD<sup>+</sup> depletion in cancer cells[J]. Cell Rep, 2018, 25(11): 3047-3058.
- [12] Fantin VR, St-Pierre J, Leder P. Attenuation of LDH-a expression uncovers a link between glycolysis, mitochondrial physiology, and tumor maintenance[J]. Cancer Cell, 2006, 9(6): 425-434.
- [13] Vander Heiden MG. Targeting cancer metabolism: a therapeutic window opens[J]. Nat Rev Drug Discov, 2011, 10(9): 671-684.
- [14] 赵雪可,李威,刘文,等. LDHA 对乳腺癌演变的影响及 SUMO 化修饰[J].临床与实验病理学杂志,2022,38(4):427-431.

  Zhao XK, Li W, Liu W, et al. Effect of LDHA on the evolution of breast cancer and SUMOylation modification[J]. Chin J Clin Exp Pathol, 2022, 38(4): 427-431.

- [15] Eilertsen M, Andersen S, Al-Saad S, et al. Monocarboxylate transporters 1-4 in NSCLC: MCT1 is an independent prognostic marker for survival[J]. PLoS One, 2014, 9(9): e105038.
- [16] 张倩,唐镇.基于 Oncomine 数据库分析 *SLC16A1* 基因在口腔癌等头颈癌中的表达和预后[J].南昌大学学报(医学版),2018,58(4):10-14,43.
  - Zhang Q, Tang Z. *SLC16A1* gene expression and prognosis in oral cancer and other head and neck cancer; an analysis based on oncomine database[J]. J Nanchang Univ Med Sci, 2018, 58(4); 10–14, 43.
- [17] Zhang L, Song ZS, Wang ZS, et al. High expression of SLC16A1 as a biomarker to predict poor prognosis of urological cancers[J]. Front Oncol, 2021, 11: 706883.
- [18] Yuan C, Zhang J, Lou JJ, et al. Comprehensive Analysis of Monocarboxylate Transporter 4 (MCT4) expression in breast cancer prognosis and immune infiltration via integrated bioinformatics analysis [J]. Bioengineered, 2021, 12(1): 3850-3863.
- [19] Johnson JM, Cotzia P, Fratamico R, et al. MCT1 in invasive ductal carcinoma; monocarboxylate metabolism and aggressive breast cancer [J]. Front Cell Dev Biol, 2017, 5; 27.
- [20] Khan A, Valli E, Lam H, et al. Targeting metabolic activity in high-risk neuroblastoma through Monocarboxylate Transporter 1 (MCT1) inhibition[J]. Oncogene, 2020, 39(17): 3555-3570.
- [21] Hou L, Zhao Y, Song GQ, et al. Interfering cellular lactate homeostasis overcomes Taxol resistance of breast cancer cells through the microRNA-124-mediated lactate transporter (MCT1) inhibition[J]. Cancer Cell Int, 2019, 19: 193.
- [22] Sun Q, Hu LL, Fu Q. MCT4 promotes cell proliferation and invasion of castration-resistant prostate cancer PC-3 cell line [J].

- EXCLI J, 2019, 18: 187-194.
- [23] Dong LQ, Lu DY, Chen R, et al. Proteogenomic characterization identifies clinically relevant subgroups of intrahepatic cholangiocarcinoma[J]. Cancer Cell, 2022, 40(1): 70-87.
- [24] Xue L, Liu JY, Xie JH, et al. Prognostic value of SLC16A3 (MCT4) in lung adenocarcinoma and its clinical significance [J]. Int J Gen Med, 2021, 14: 8413-8425.
- [25] Zhao Y, Zhao B, Yan WH, et al. Integrative analysis identified MCT4 as an independent prognostic factor for bladder cancer [J]. Front Oncol, 2021, 11: 704857.
- [26] Mukai Y, Yamaguchi A, Sakuma T, et al. Involvement of SLC16A1/MCT1 and SLC16A3/MCT4 in l-lactate transport in the hepatocellular carcinoma cell line [J]. Biopharm Drug Dispos, 2022, 43(5): 183-191.
- [27] Tomczak K, Czerwińska P, Wiznerowicz M. The Cancer Genome Atlas (TCGA): an immeasurable source of knowledge[J]. Contemp Oncol, 2015, 19(1A): A68-A77.
- [28] Zhang Z, Li H, Jiang S, et al. A survey and evaluation of Webbased tools/databases for variant analysis of TCGA data [J]. Brief Bioinform, 2019, 20(4): 1524-1541.
- [29] 张曼,周兰柱,孙哲,等.基于 TCGA 数据库分析 POSTN 在头颈 部鳞癌中的表达和临床意义[J].中华全科医学,2022,20(6): 1066-1070.
  - Zhang M, Zhou LZ, Sun Z, et al. Analysis of the expression and clinical significance of POSTN in head and neck squamous carcinoma based on TCGA database [J]. Chin J Gen Pract, 2022, 20(6): 1066-1070.

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#### (上接第347页)

- [32] Gu JH, Jiang TA. Ultrasound radiomics in personalized breast management: current status and future prospects [J]. Front Oncol, 2022, 12: 963612.
- [33] Zhou BY, Wang LF, Yin HH, et al. Decoding the molecular subtypes of breast cancer seen on multimodal ultrasound images using an assembled convolutional neural network model: a prospective and multicentre study[J]. EBioMedicine, 2021, 74: 103684.
- [34] Jiang M, Zhang D, Tang SC, et al. Deep learning with convolutional neural network in the assessment of breast cancer molecular subtypes based on US images: a multicenter retrospective study[J]. Eur Radiol, 2021, 31(6): 3673-3682.
- [35] Liu Y, Wang Y, Wang YX, et al. Early prediction of treatment response to neoadjuvant chemotherapy based on longitudinal ultrasound images of HER2-positive breast cancer patients by Siamese multitask network: a multicentre, retrospective cohort study[J]. EClinicalMedicine, 2022, 52: 101562.
- [36] Jiang M, Li CL, Luo XM, et al. Ultrasound-based deep learning radiomics in the assessment of pathological complete response to neoadjuvant chemotherapy in locally advanced breast cancer[J]. Eur J Cancer, 2021, 147; 95-105.

- [37] Gu JH, Tong T, He C, et al. Deep learning radiomics of ultrasonography can predict response to neoadjuvant chemotherapy in breast cancer at an early stage of treatment; a prospective study[J]. Eur Radiol, 2022, 32(3): 2099-2109.
- [38] Lee YW, Huang CS, Shih CC, et al. Axillary lymph node metastasis status prediction of early-stage breast cancer using convolutional neural networks [J]. Comput Biol Med, 2021, 130: 104206.
- [39] Zhou LQ, Wu XL, Huang SY, et al. Lymph node metastasis prediction from primary breast cancer US images using deep learning [J]. Radiology, 2020, 294(1): 19-28.
- [40] Tahmasebi A, Qu EZ, Sevrukov A, et al. Assessment of axillary lymph nodes for metastasis on ultrasound using artificial intelligence [J]. Ultrason Imaging, 2021, 43(6): 329-336.
- [41] Gu JH, Tong T, Xu D, et al. Deep learning radiomics of ultrasonography for comprehensively predicting tumor and axillary lymph node status after neoadjuvant chemotherapy in breast cancer patients: a multicenter study[J]. Cancer, 2023, 129(3): 356-366.

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